

SYNTHESIS OF [³²P] LABELLED 1-O-ALKYL-2-DESOXY-2-AMINO-*SN*-GLYCERO-3- PHOSPHOCHOLINES

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SUMMARY

The syntheses of N-substituted 1-O-alkyl-2-desoxy-2-amino-*sn*-glycero-3-[³²P]phosphocholines were performed in four steps starting from [³²P] POCl₃ and the corresponding 1-O-alkyl-2-amino-propane-3-ols in 5-7 % total yield.

KEY WORDS

[³²P]etherphospholipids; 1-O-alkyl-2-desoxy-2-amino-*sn*-glycero-3-[³²P]phosphocholines

INTRODUCTION

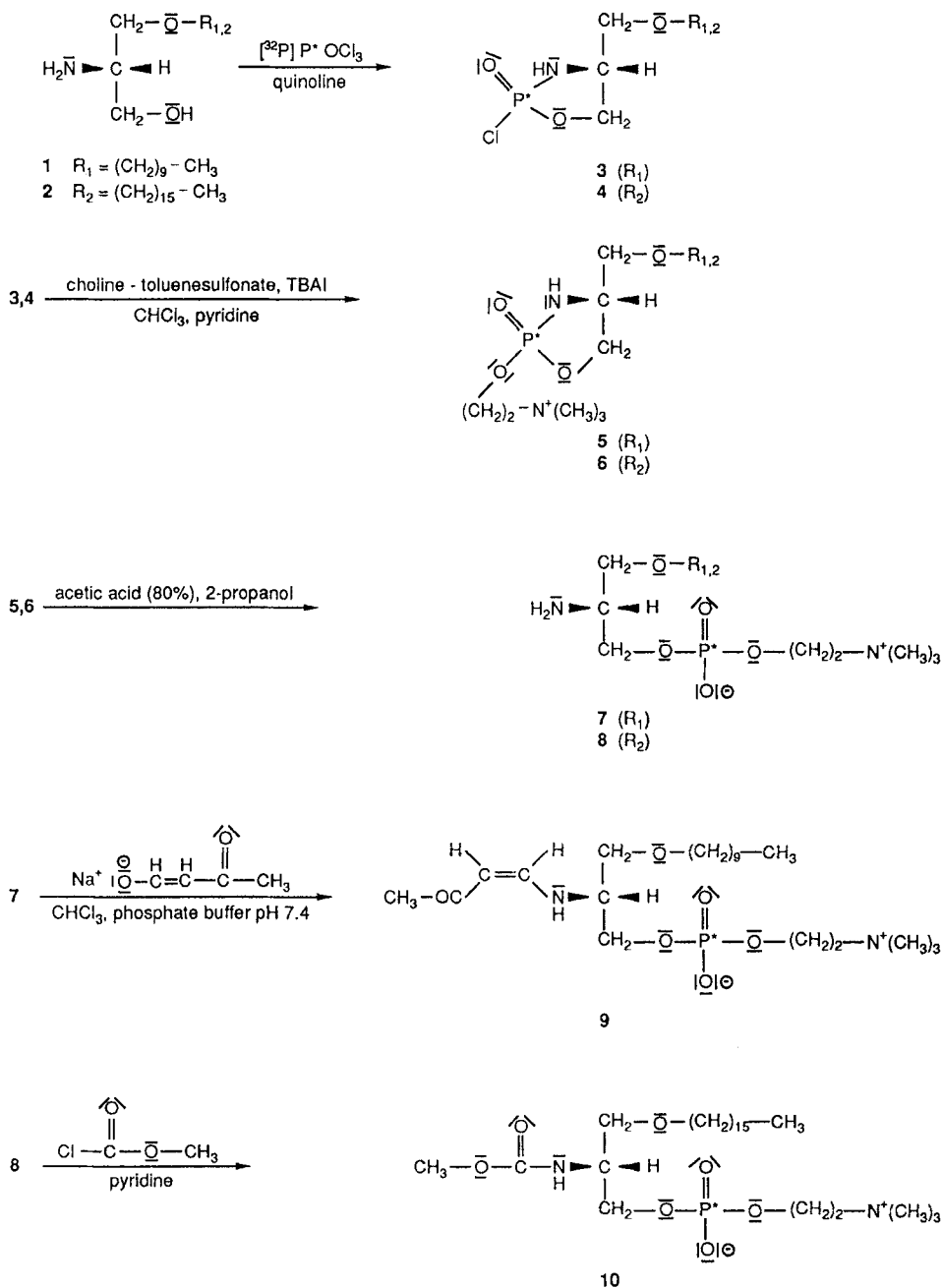
Derivatives of 1-O-alkyl-2-desoxy-2-amino-*sn*-glycero-3-phosphocholine are potent inhibitors of phospholipase A₂ [1, 2] and potential ligands of the receptor of the platelet activating factor (PAF). Isotopically labelled, 2-amino-etherphospholipids therefore are of potential value in investigations involving interactions of phospholipids with cellular membranes as well as in metabolic studies.

In this report we describe the preparation of [³²P] labelled 2-desoxy-2-amino-*sn*-glycero-etherphospholipids utilizing [³²P] POCl₃ to introduce the radioactive label and 4-substituted 2-chloro-2-oxo-1,3,2-oxaza-[³²P]phospholanes as versatile intermediates.

RESULTS AND DISCUSSION

The synthetic route to labelled 1-O-alkyl-2-desoxy-2-amino-*sn*-glycero-3-phosphocholines is outlined in the scheme. Preparation of [³²P] labelled 2-desoxy-2-amino-lysophospholipids was carried out according to the synthetic sequence for unlabelled 2-amino-phospholipids published by us previously [3].

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SCHEME

Equilibration of [^{32}P] phosphoric acid with phosphorus oxychloride yielded isotopically labelled POCl_3 ; reaction with the corresponding 1-O-alkyl-2-amino-propane-3-ol (**1**, **2**) gave the labelled oxazaphospho-
 lane intermediates (**3**, **4**). The choline group was introduced by nucleophilic exchange of the chlorine, acid

hydrolysis of (5), (6) opened the ring to give the desired 1-*O*-alkyl-2-desoxy-2-amino-*sn*-glycero-[³²P]-phosphocholines (7) and (8). The total yield of the three consecutive steps was lower starting from the long-chain aminoalcohol (2) (12% vs. 32%) suggesting a decreased reactivity along with increasing carbon chain length. Conversion of the 2-amino-lysophospholipids (7) and (8) with the sodium salt of 1-hydroxy-but-1-en-3-on or with methylchloroformate and purification by chromatography on thin layer plates provided the labelled vinyllogous amide (9) and the carbamic-acid-methylester (10) with a specific activity of approximately 720 MBq/mmol and a radiochemical purity of 95-96%. The specific radioactivity of the phospholipids was 0.78 of the theoretical value, a result which can be explained by incomplete exchange with [³²P]phosphoric acid.

MATERIALS

Tetrabutyl ammonium iodide was purchased from Fluka (Buchs, Switzerland), phosphorus oxychloride and *p*-toluenesulfonic acid from Aldrich (Steinheim, Germany). Chloroform and methanol were obtained from J. T. Baker (Deventer, Holland) and distilled from P₂O₅ and from Mg prior to use. Silica gel (grade 60, 70-230 mesh) for column chromatography and 2-propanol was from Machery-Nagel (Düren, Germany). TLC plates (0.5 mm, F 254) were from E. Merck (Darmstadt, Germany) and preeluted with methanol. [³²P]phosphoric acid (314-337 TBq/mmol) was from Du Pont de Nemours Deutschland GmbH (Bad Homburg).

METHODS

Thin-layer chromatography (TLC) and column chromatography were performed using a mixture of chloroform/methanol/water (65:45:8, v/v) as mobile phase. Phospholipids were detected with "Phospray" (Supelco, Bad Homburg, Germany). Mass spectra were obtained using a MAT 311 A mass spectrometer (Varian, Bremen, Germany) equipped with a FAB ion gun (Xe, 6 KV, ion current 1 mA) from Ion Tech (Teddington, U. K.) and glycerol as matrix. ¹H-NMR-spectra were recorded at 250 MHz (WM-250, Bruker Physik AG, Karlsruhe, Germany); tetramethyl silane was used as internal reference. Spectra were run in acetonitrile-D₃ or in CDCl₃/methanol-D₄, 2:1 (v/v). Multiplicities are reported as singlet (s), doublet (d), triplet (t) or multiplet (m).

[³²P] phosphorus oxychloride

Labelled phosphorus oxychloride was obtained applying the method of Keenan et al. [4].

[³²P] phosphoric acid (370 MBq, 314-337 TBq/mmol) was mixed with freshly distilled phosphorus oxychloride (30 μl, 0.32 mmol) to give a theoretical specific activity of 1.12 GBq/mmol (calculated for 100% conversion to POCl₃) and stirred for 24 h at 107°C in a screwed vial.

1-O-decyl-2-desoxy-2-amino-*sn*-glycero-[³²P]phosphocholine (7) and 1-O-hexadecyl-2-desoxy-2-amino-*sn*-glycero-3-[³²P]phosphocholine (8)

To 12 μ l (129 μ mol) of [³²P] phosphorus oxychloride dissolved in chloroform, a solution of 30 mg (130 μ mol) of 1-O-decyl-2-amino-propane-3-ol (**1**) or 41 mg (130 μ mol) 1-O-hexadecyl-2-amino-propane-3-ol (**2**) and 35 μ l (290 μ mol) quinoline was added dropwise at 4°C. The mixture was allowed to warm up at room temperature, then stirred at 55°C for 16 h.

The resulting solution of 1-O-decyl-2,3-(2'-chloro-2'-oxo-1',3',2')-oxaza[³²P]phospholane (**3**) or 1-O-hexadecyl-2,3-(2'-chloro-2'-oxo-1',3',2')-oxaza[³²P]phospholane (**4**) was cooled to 12°C and 55mg (0.2 mmol) choline tosylate in 0.3 ml pyridine was added. After stirring for 24 h at 55°C, the solvents were removed *in vacuo*, and the residue containing the crude oxazaphospholanes (**5**) or (**6**) was redissolved in 1 ml of 2-propanol/acetic acid (80%) 3:2 (v/v). Hydrolysis was carried out by stirring at 50°C for 30 min and at room temperature for additional 2 h. The solvents were removed by distillation *in vacuo* at under 40°C and silica gel chromatography afforded 16.3 mg (41 μ mol, 31%) of (**7**) and 7.2 mg (15 μ mol, 12%) of (**8**); MS (FAB; glycerol; pos. mode): $m/z = 397$ [M+H]⁺ (**7**) and $m/z = 481$ [M+H]⁺ (**8**).

1-O-decyl-2-desoxy-2-(1'-amino-but-1'-en-3'-on)-*sn*-glycero-3-phosphocholine (9)

To 16.3 mg (41 μ mol) of (**7**), dissolved in 2 ml of a biphasic mixture (1:1, v/v) of chloroform/phosphate buffer (20 mM, pH 7.4) 200 mg (1.85 mmol) of the sodium salt of 1-hydroxy-but-1-en-3-on [5] were added and the solution stirred for 24 h at room temperature. The solvents were removed by distillation *in vacuo* and the residue purified by thin layer chromatography yielding 2.8 mg (6 μ mol, 15%) of (**9**) (cis configuration, specific activity 724 MBq/mmol)

MS (FAB; glycerol; pos. mode): $m/z = 465$ [M+H]⁺

¹H-NMR (CDCl₃/D₄-methanol, 2:1) δ ppm:

6.8 (1H, d, -CH=CH-C=O), 4.95 (1H, d, -CH=CH-C=O), 4.2 (1H, m, *sn*-2-CH), 3.95 (2H, m, CH₂-CH₂-N⁺(CH₃)₃), 3.9-3.8 (2H, t, *sn*-3-CH₂), 3.6 (2H, m, -CH₂-N⁺(CH₃)₃), 3.45 (2H, m, *sn*-1-CH₂-), 3.3 (3H, s, O=C-CH₃), 3.2 (9H, s, -N⁺(CH₃)₃), 1.5 (2H, m, -CH₂-(CH₂)₈-CH₃), 1.25 (16H, m, -CH₂-(CH₂)₈-CH₃), 0.85 (3H, t, -(CH₂)₉-CH₃).

1-O-hexadecyl-2-desoxy-2-amino-carbamic-acid-methylester-*sn*-glycero-3-phosphocholine (10)

To a solution of 7.2 mg (15 μ mol) (**8**) in chloroform/pyridine (3 ml, 5:1), 10 μ l (130 μ mol) of methylchloroformate were added dropwise at 0°C; the reaction mixture was allowed to warm up to room temperature and stirred for another 4 h. Evaporation *in vacuo* afforded crude (**10**) which was subjected to thin layer chromatography to give 4.8 mg (9 μ mol, 60%) of (**10**) (specific activity 715 MBq/mmol)

MS (FAB; glycerol; pos. mode): $m/z = 523$ [M+H]⁺.

¹H-NMR (CD₃-C≡N) δ ppm:

4.2 (1H, m, *sn*-2-CH), 3.9 (2H, m, CH₂-CH₂-N⁺(CH₃)₃), 3.9-3.8 (2H, m, *sn*-3-CH₂), 3.6 (3H, s, -O-CH₃), 3.55 (2H, m, -CH₂-N⁺(CH₃)₃), 3.45 (2H, m, *sn*-1-CH₂-), 3.2 (9H, s, -N⁺(CH₃)₃), 1.5 (2H, m, -CH₂-(CH₂)₁₄-CH₃), 1.25 (28H, m, -CH₂-(CH₂)₁₄-CH₃), 0.85 (3H, t, -(CH₂)₁₅-CH₃).

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